Analytical Evidence Report

HPLC-UV results for the submitted KLOW Blend sample

REPORT NUMBER
VDIC-KLOW-80-110326

GENERATED
November 09, 2026

KLOW Blend

Sample vial 2, injection 1 | Client report VDIC-KLOW-80-110326

Results and chromatograms

Lot # 961026 | Client: Accutide Research

Laboratory Analysis: LAB-CA-02

Laboratory identity and applicable credentials are maintained by Testides and are available to regulatory authorities where required by law.

VIAL PHOTOGRAPH	SAMPLE INFORMATION		ANALYSIS		RUN DETAILS	
	Peptide:	**KLOW Blend**	Method:	**HPLC-UV**	Run date:	**Nov 05, 2026**
	Client ID:	**VDIC-KLOW-80-110326**	Channel:	**214 nm**	Acquired:	**16:56:26 UTC**
	Label claim:	**80 mg**	Integration:	**Laboratory**	Run time:	**25 min**
	Injection:	**10 uL, 214 nm**	Identity:	**Conforms**	Laboratory:	**LAB-CA-02**

RESULTS AT A GLANCE

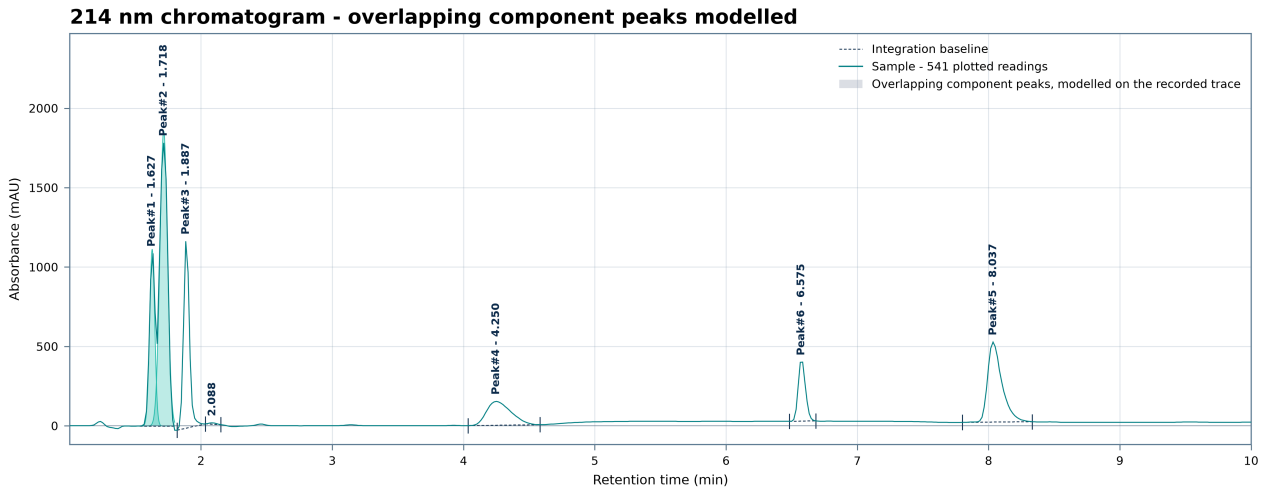
PROJECTED AMOUNT	HPLC PURITY	CONTENT VS. LABEL	IDENTITY
76.51 mg*	**99.96%**	**95.6%**	**Conforms**
SUM OF COMPONENTS; SOME EXTRAPOLATED	NAMED COMPONENTS, 214 NM*	76.51 MG OF 80 MG CLAIMED	PEAK IDENTITY CONFIRMED**

GHK-CU	KPV	BPC-157	TB-500
53.00 mg*	**5.65 mg†**	**8.81 mg†**	**9.06 mg†**
106.0% OF 50 MG LABEL; EXTRAPOLATED	56.5% OF 10 MG LABEL	88.1% OF 10 MG LABEL	90.6% OF 10 MG LABEL

ENDOTOXINS
1.0 EU/mL
BELOW LIMIT (≤ 1 EU/ML)

CHROMATOGRAM

Every plotted reading as recorded - overlapping component peaks drawn as peak shapes fitted to the trace



HPLC PURITY (214 NM)

Named components	Integrated peaks	Smallest peak integrated	Recovered chromatogram
99.96%	**7**	**0.19%**	**±0.77%**
Laboratory integration	Counted in the total	Of total area	Area vs. laboratory table

Purity is the component peaks' share of the total peak area the laboratory integrated at 214 nm. The smallest peak it integrated was 0.19% of the total; the laboratory does not state a reporting threshold, so peaks below the ones it integrated are not counted. Integrating the recovered chromatogram reproduces its component area to within 0.77%. This result rests on a single injection.

The chromatogram is recovered point for point from the laboratory's report: 541 plotted readings from 1.00 to 10.00 min, joined as drawn; nothing is smoothed or resampled.

* HPLC purity is the component peaks' share of all peak area the laboratory integrated; every peak is itemised on page 3.

** Identity conforms. GHK-Cu, KPV, BPC-157, TB-500 are identified by elution order; no reference on the sample's method.

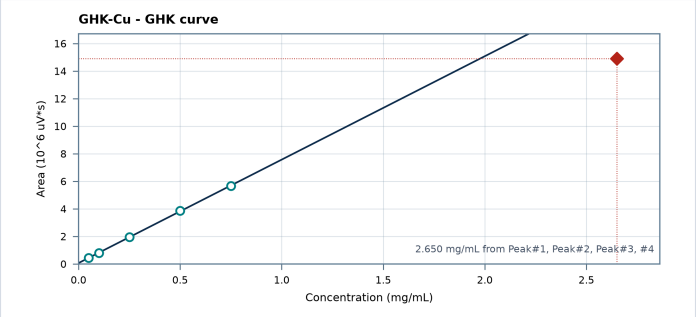
KLOW Blend

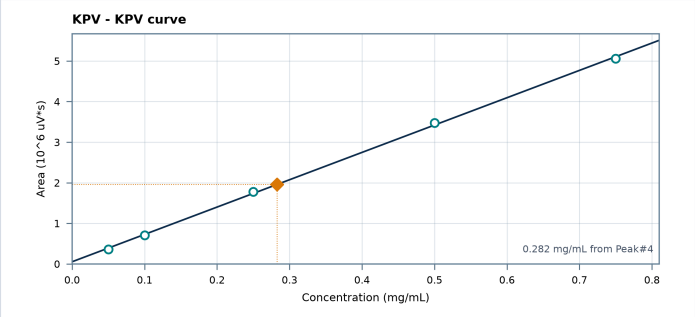
Sample vial 2, injection 1 | Client report VDIC-KLOW-80-110326

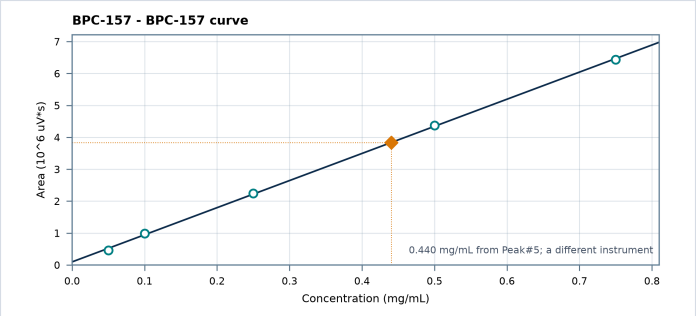
Reference comparison and calculation

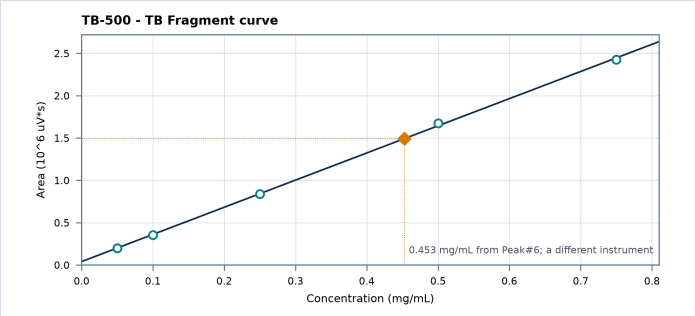
Lot # 961026 | Client: Accutide Research

CONCENTRATION CALIBRATION - 214 NM (LABORATORY VALIDATION CURVES)





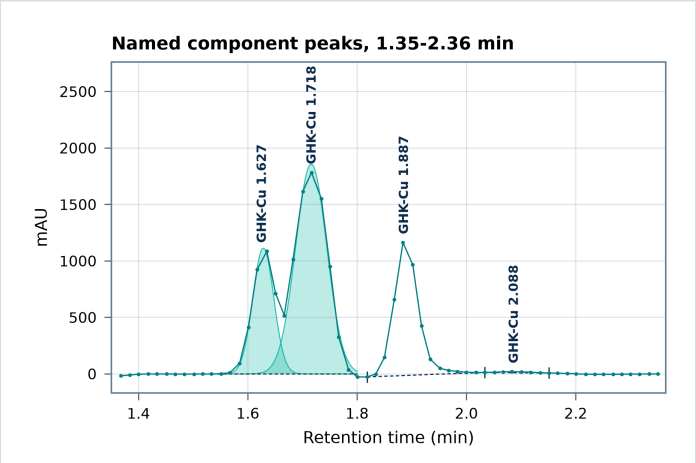


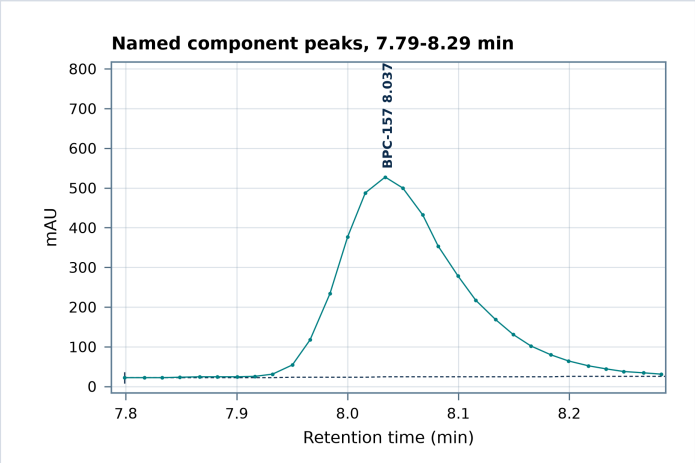


Component	Peaks	Area %	mg/mL injected	Laboratory curve	Amount	Label share	Content
GHK-Cu	Peak#1, Peak#2,...	60.65%	2.6499 *	GHK	53.00 mg	50 mg	106.0%
KPV	Peak#4	10.57%	0.2825	KPV	5.65 mg	10 mg	56.5%
BPC-157	Peak#5	20.72%	0.4403	BPC-157	8.81 mg	10 mg	88.1%
TB-500	Peak#6	8.06%	0.4528	TB Fragment	9.06 mg	10 mg	90.6%

† The vial is reconstituted in 2 mL of diluent and diluted x10, which puts the component present in the smallest amount at the working 0.5 mg/mL. Label shares follow the blend's recorded composition: GHK-Cu 50 mg, KPV 10 mg, BPC-157 10 mg, TB-500 10 mg. Total 76.51 mg. * Outside the range of the laboratory's curve; the reading is extrapolated. Curves validated on a different instrument are applied to this injection.

NAMED COMPONENT PEAKS





Native plotted readings, nothing smoothed. Overlapping component peaks are drawn and read as peak shapes fitted to the readings, on one baseline under the group, not the laboratory's valley-to-valley integration, which leaves the area beneath the valley in neither peak. Purity is the laboratory's.
Each component is confirmed by retention time against its reference where one is on the sample's method, otherwise by elution order; a component can show as more than one peak.

KLOW Blend

Sample vial 2, injection 1 | Client report VDIC-KLOW-80-110326

Definitions, calculation detail, and scope

Lot # 961026 | Client: Accutide Research

HOW TO READ THIS REPORT

Start with the result on page 1. Pages 2 and 3 show the curve, comparison, and peaks behind it.

01 **Recovered chromatogram**

The laboratory's trace, read back point for point from its report.

02 **Integrated peak table**

Every peak the laboratory integrated, and which one is the analyte.

03 **Validation curve**

Converts the peak area to a concentration for this analyte.

WHAT THE RECOVERED CHROMATOGRAM ADDS

How the chromatogram on this report was obtained

How it was recovered

The laboratory's data system draws the chromatogram in its report as vector data. Every vertex it drew was read back - 541 readings - and both axes were calibrated from the report's own tick marks. Integrating the recovered trace over the laboratory's baselines and peak marks reproduces the area in its peak table to within 0.77%.

What it can and cannot show

It shows every peak, shoulder and baseline the laboratory integrated, at the resolution the laboratory printed. It carries a single wavelength and no UV spectrum, so it cannot reveal a co-eluting compound the way diode-array data can.

Scope: the recovered chromatogram reproduces the laboratory's report. It does not re-measure the sample.

HOW THE PURITY RESULT WAS MEASURED

214 nm area percent, from the laboratory's own integration

The laboratory integrated each peak to its own baseline and named the analyte's peak in its processing method. Purity is that peak's share of the total integrated area; the shares below are recomputed from the areas, and each peak's area is checked against the recovered chromatogram.

Purity (%) = assigned peak area / total integrated peak area x 100

Peak	RT (min)	Area (uV*s)	Area %	Tailing	Plates	Role	Recovered
1	1.627	1,915,270	10.35	0.94	6,384	Peak#1 - GHK-Cu	1,917,019 (+0.09%)
2	1.718	5,574,279	30.11	1.11	4,107	Peak#2 - GHK-Cu	5,617,397 (+0.77%)
3	1.887	3,703,685	20.01	1.09	6,602	Peak#3 - GHK-Cu	3,730,713 (+0.73%)
4	2.088	34,388	0.19	1.06	5,398	GHK-Cu	-
5	4.250	1,956,849	10.57	1.32	2,254	Peak#4 - KPV	1,953,514 (-0.17%)
6	6.575	1,491,816	8.06	1.14	57,184	Peak#6 - TB-500	1,498,220 (+0.43%)
7	8.037	3,836,831	20.72	1.49	25,938	Peak#5 - BPC-157	3,823,839 (-0.34%)

Integrated peaks 7 | smallest 0.19% of total | injections 1

Area percent assumes every peak absorbs at 214 nm like the main peak; the analyte's peak is the one the laboratory's method names. Recovered areas are compared for the analyte's peaks of 1% or more.

WHAT THIS REPORT CAN AND CANNOT TELL YOU

The report explains the measurement; it does not decide how a material should be used

Observed evidence

The laboratory's chromatogram, its integration, and its validation curve.

Calculated results

Area-percent purity, concentration, projected amount, and calibration trace.

Method context

Retention time against the reference standard or the report's other vials, where on the same method.

Safety and regulatory status

No conclusion on safety, efficacy, fitness for use, or regulatory status.

This analytical record does not authorize or endorse manufacture, sale, marketing, distribution, or use of the material.

KLOW Blend

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Endotoxin result and readings

Lot # 961026 | Client: Accutide Research

ENDOTOXIN RESULT

RESULT	THRESHOLD	METHOD	CONTROLS
PASS	**≤ 1 EU/mL**	**Gel-clot LAL**	**Passed**
NEGATIVE BY THE LABORATORY	AT OR BELOW PASSES	USP <85>	POSITIVE AND NEGATIVE

At or below the 1 EU/mL threshold

BACTERIAL ENDOTOXINS — HOW THE RESULT WAS MEASURED

Measured value	**1.0 EU/mL**	Laboratory result	**Negative**
Mean endpoint	**1/8**	Lysate sensitivity (λ)	**0.125 EU/mL**
Lysate	**-**	Lysate lot	**DC0423**
Dilutions	**1/2 to 1/32, two-fold**	Replicates	**2**
Controls	**Passed**	Control standard	**-**
Sample	**Submitted vial**	Incubation	**-**
Tested	**Nov 05, 2026**	Approved	**Nov 09, 2026**

Gel-clot readings: each replicate at each two-fold dilution of the sample solution, + firm gel, - no gel

Replicate	1/2	1/4	1/8	1/16	1/32	Endpoint
1	+	+	+	-	-	1/8
2	+	+	+	-	-	1/8

How the concentration is calculated

Endotoxin concentration = lysate sensitivity / mean endpoint = 0.125 EU/mL / 1/8 = 1.0 EU/mL. The mean endpoint is the geometric mean of the replicates' endpoints (the most dilute tube that gels).

Laboratory worksheet, tested Nov 05, 2026, approved Nov 09, 2026. Lysate: lot DC0423. The positive control gelled and the negative control did not, as required.

ABOUT THE TEST

The bacterial endotoxins test detects endotoxin from Gram-negative bacteria with limulus amebocyte lysate (LAL), which forms a firm gel when endotoxin at or above the lysate's labelled sensitivity is present. The laboratory tested the sample solution in a two-fold dilution series (1/2 to 1/32) in 2 replicates; a tube that holds a firm gel when inverted is positive.

Each replicate's endpoint is the most dilute tube that gels, and the endotoxin concentration of the solution is the labelled sensitivity divided by the geometric mean of the endpoints. For the series to be valid a positive control (control standard endotoxin) must gel and a negative control (endotoxin-free water) must not.

The result is PASS when the laboratory reports the sample negative and FAIL when it reports it positive; the threshold it is stated against is ≤ 1 EU/mL, inclusive.